

Total Synthesis of (–)-Reveromycin A

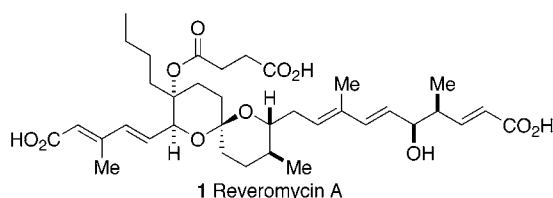
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ABSTRACT



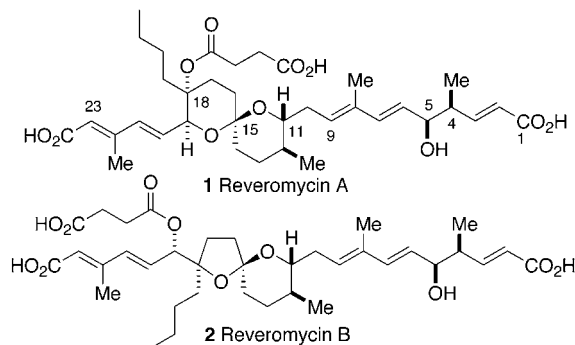
The asymmetric total synthesis of (–)-reveromycin A is described. The key steps involved a Lewis acid catalyzed inverse electron demand hetero-Diels–Alder reaction followed by hydroboration/oxidation to afford the spiroketal core **4** in a highly stereoselective manner and introduction of the C18 hemisuccinate by high-pressure acylation.

The reveromycins A (**1**) and B (**2**) are members of a family of compounds isolated from the soil actinomycete *Streptomyces* sp.¹ Reveromycin A (**1**) is a potent inhibitor (IC_{50} 0.7 $\mu\text{g mL}^{-1}$) of the mitogenic activity of epidermal growth factor (EGF) in a mouse keratinocyte. In addition, compound **1** exhibits antiproliferative activity against human tumor cell lines (IC_{50} = 1.3–2.0 $\mu\text{g mL}^{-1}$), as well as antifungal activity (MIC = 2.0 $\mu\text{g mL}^{-1}$, pH 3).^{1b} Recently, reveromycin A (**1**) has been identified as a specific inhibitor of *Saccharomyces cerevisiae* isoleucyl-tRNA synthetase (IleRS) using yeast genetics and biochemical studies.²

So far, only one total synthesis of **1** has been reported,³ and several approaches to the 6,6-spiroketal core have been described,^{4–6} while three total syntheses of reveromycin B (**2**) have been completed.^{7–9}

We now report the total synthesis of (–)-reveromycin A (**1**) using a hetero-Diels–Alder (HDA) strategy to construct the challenging spiroketal moiety of this molecule.⁶

It is known that the reveromycin A (**1**)-type 6,6-spiroketal core **I** can undergo acid-catalyzed isomerization to afford the spiroisomer **II** as the result of an unfavorable steric interaction involving the axial C19 side chain in **I** that is alleviated in **II**, albeit at the cost of an anomeric effect (Figure 1). Thus, the energy difference between the 6,6-



- (1) (a) Osada, H.; Koshino, H.; Isono, K.; Takahashi, H.; Kawanishi, G. *J. Antibiot.* **1991**, *44*, 259. (b) Takahashi, H.; Osada, H.; Koshino, H.; Kudo, T.; Amano, S.; Shimizu, S.; Yoshihama, M.; Isono, K. *J. Antibiot.* **1992**, *45*, 1409. (c) Takahashi, T.; Osada, H.; Koshino, H.; Sasaki, M.; Onose, R.; Nakakoshi, M.; Yoshihama, M.; Isono, K. *J. Antibiot.* **1992**, *45*, 1414. (d) Koshino, H.; Takahashi, H.; Osada, H.; Isono, K. *J. Antibiot.* **1992**, *45*, 1420.
- (2) Miyamoto, Y.; Machida, K.; Mizunuma, M.; Emoto, Y.; Sato, N.; Miyahara, K.; Hirata, D.; Usui, T.; Takahashi, H.; Osada, H.; Miyakawa, T. *J. Biol. Chem.* **2002**, *277*, 28810.
- (3) Shimizu, T.; Masuda, T.; Hiramoto, K.; Nakata, T. *Org. Lett.* **2000**, *2*, 2153.
- (4) Shimizu, T.; Kobayashi, R.; Osako, K.; Osada, H.; Nakata, T. *Tetrahedron Lett.* **1996**, *37*, 6755.
- (5) Drouet, K. E.; Ling, T.; Tran, H. V.; Theodorakis, E. A. *Org. Lett.* **2000**, *2*, 207.
- (6) El Sous, M.; Rizzacasa, M. A. *Tetrahedron Lett.* **2000**, *41*, 8591.
- (7) (a) Drouet, K. E.; Theodorakis, E. A. *J. Am. Chem. Soc.* **1999**, *121*, 456. (b) Drouet, K. E.; Theodorakis, E. A. *Eur. J. Chem.* **2000**, *6*, 1987.
- (8) Masuda, T.; Osako, K.; Shimizu, T.; Nakata, T. *Org. Lett.* **1999**, *1*, 941.
- (9) (a) Cuzzupe, A. N.; Hutton, C. A.; Lilly, M. J.; Mann, R. K.; Rizzacasa, M. A.; Zammit, S. C. *Org. Lett.* **2000**, *2*, 191. (b) Cuzzupe, A. N.; Hutton, C. A.; Lilly, M. J.; Mann, R. K.; McRae, K. J.; Rizzacasa, M. A.; Zammit, S. C. *J. Org. Chem.* **2001**, *66*, 2382.

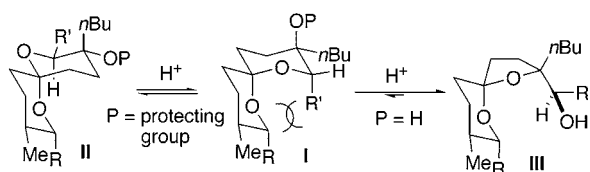
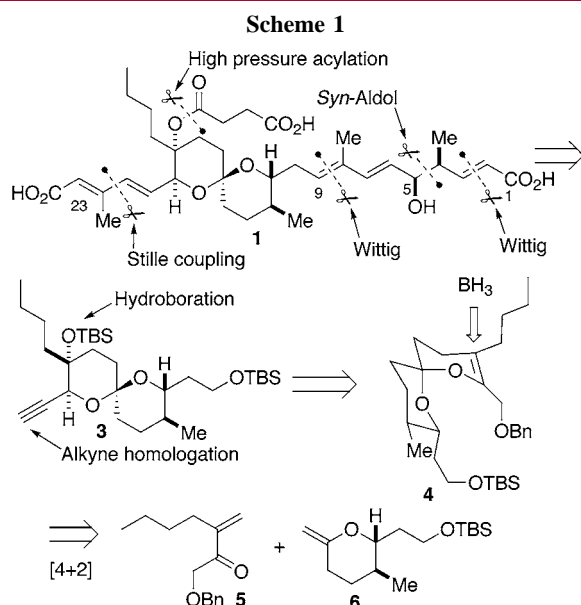


Figure 1.

spiroketals **I** and **II** is small. In addition, when the C18 alcohol is free, acid-induced isomerization provides the more stable reveromycin B (**2**) 5,6-spiroketal core **III** exclusively. This energy profile has been observed in the reported thermodynamic approaches (i.e., cyclization of a “dihydroxy-ketone” precursor) to the 6,6-spiroketal of **1**, which provided mixtures of spiroisomers,^{3–5} and degradative^{4,10} and synthetic studies^{6,7,9b,11} have demonstrated the strong preference for the reveromycin B-type 5,6-spiroketal **III**.

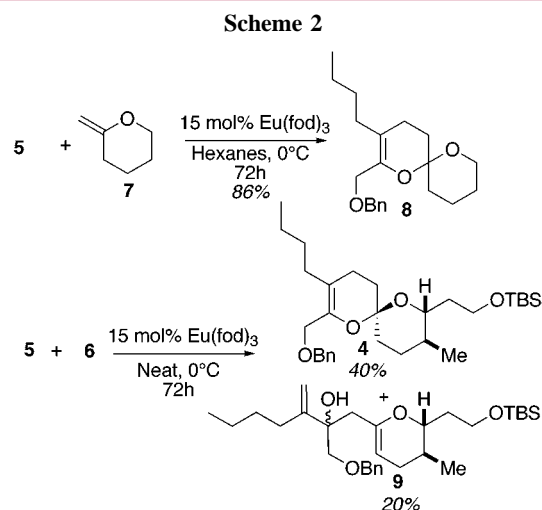
In light of the above results, we envisioned that a kinetic approach to the reveromycin A spiroketal may circumvent the thermodynamic dilemma cited above. As shown in Scheme 1, the indicated retrosynthetic disconnections of **1**



lead to the intermediate 6,6-spiroketal **3**. This could be obtained from unsaturated spiroketal **4** by regio- and stereoselective hydroboration from the direction shown followed by alkyne homologation. Key spiroketal **4** could be obtained by an inverse electron demand HDA reaction¹² between the diene **5**⁶ and the chiral methylene pyran **6**.⁹ This critical

reaction should set the stereochemistry at the spiro center by an axial approach of the carbonyl oxygen in the HDA transition state.⁹ The stereochemistry at C18 and C19 is then set by the hydroboration/oxidation sequence, which circumvents the thermodynamic lability of the reveromycin A type spiroketal **I**. In addition to being stereoselective and convergent, this route would also avoid the need for a large number of different protecting groups.

Earlier, we had found that the HDA reaction of isomerizable methylene pyrans with simple hetero dienes can be achieved thermally in the presence of K_2CO_3 , which effectively reduced the amount of the thermodynamically driven *exo* to *endo* isomerization of the dienophile.^{9,11} In the case of diene **5**, using this additive gave low yields of spiroketal because of the base lability of the diene.⁶ Therefore, Lewis acid promotion of the cycloaddition was surveyed using the diene and model dienophile **7**⁹ (Scheme 2). After some experimentation, we found that the HDA



reaction between **7** and diene **5** was promoted by $Eu(fod)_3$ ¹³ in hexane solvent at 0 °C to give the spiroketal **8** in excellent yield. Unfortunately, in the case of the substituted methylene pyran **6** required for the synthesis of **1**, no HDA reaction was observed in hexane solvent and only slow isomerization occurred to give the *endo* isomer of **6**. Eventually, we found that the cycloaddition proceeded smoothly when a neat mixture¹⁴ of the diene **5** and dienophile **6** was treated with 15 mol % $Eu(fod)_3$ at 0 °C. Although the reaction gave the desired spiroketal **4** as one diastereoisomer in a higher yield than that obtained using K_2CO_3 ,⁶ the byproduct **9**, resulting from an ene reaction, was also isolated as a mixture of diastereoisomers. This was not observed during the studies using the model pyran **7**. The HDA reaction between **5** and **6** was also promoted by $ZnCl_2$ in THF at 0 °C to provide the adduct **4** in a comparable yield.

(10) Ubukata, M.; Koshino, H.; Osada, H.; Isono, K. *J. Chem. Soc., Chem. Commun.* **1994**, 1877.

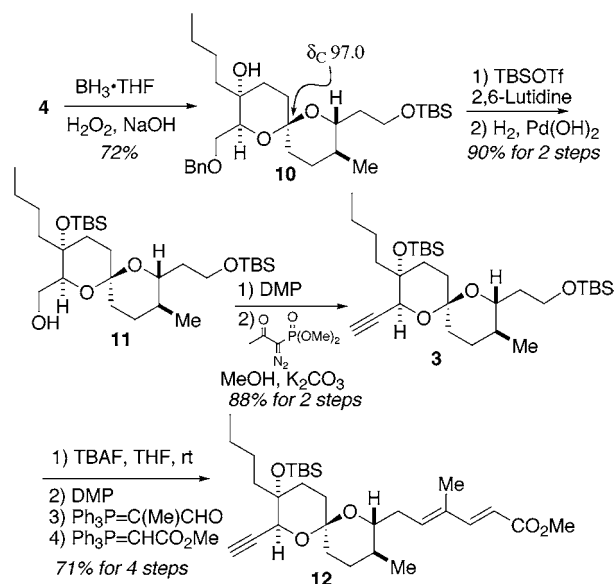
(11) McRae, K. J.; Rizzacasa, M. A. *J. Org. Chem.* **1997**, 62, 1196.

(12) For examples of the HDA synthesis of spiroketals, see: Mead, K. T.; Brewer, B. N. *Curr. Org. Chem.* **2003**, 7, 227.

(13) Bednarski, M.; Danishefsky, S. J. *J. Am. Chem. Soc.* **1983**, 105, 3716.

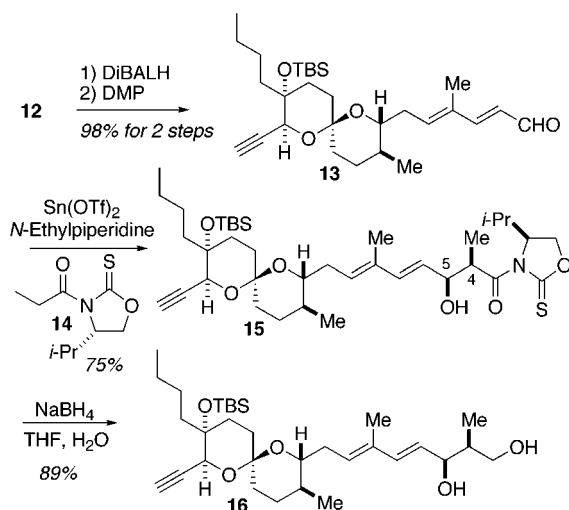
(14) Pale, P.; Bouquant, J.; Chuche, J.; Carrupt, P. A.; Vogel, P. *Tetrahedron* **1994**, 50, 8035.

Scheme 3



With the desired spiroketal in hand, we next tackled the introduction of the C18 and C19 stereocenters and C5–C10 chain extension as shown in Scheme 3. Hydroboration of spiroketal **4** followed by oxidation proceeded in good yield to afford the tertiary alcohol **10** as the only isomer. The integrity of the 6,6-spiroketal **10** was evidenced by the characteristic ^{13}C NMR chemical shift observed for the spiro carbon,^{9,11} and the stereochemistry was confirmed by its conversion into a known reveromycin A degradation product⁴ as described previously.⁶ Protection of the tertiary alcohol followed by debenzoylation afforded alcohol **11** ready for alkyne homologation. Oxidation¹⁵ of **11** and alkyne formation following the Bestmann protocol¹⁶ then gave compound **3**. Selective desilylation of the primary TBS group (TBAF, rt) followed by oxidation and two-stage Wittig extension provided the diene **12** in excellent overall yield for the four

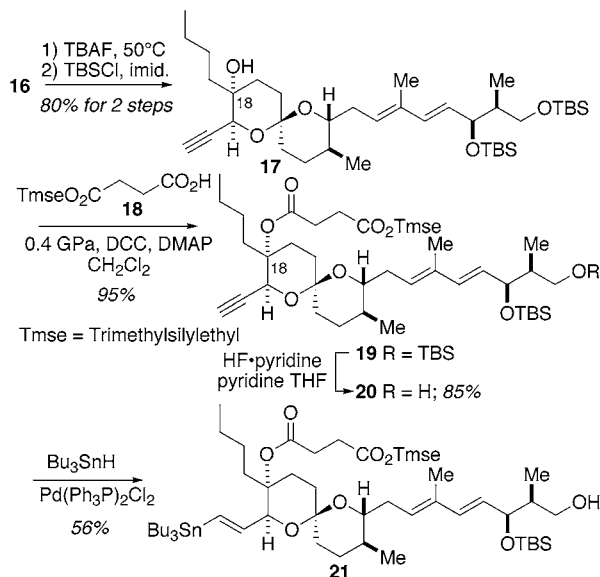
Scheme 4



steps. Reduction of **12** and subsequent oxidation provided the substrate **13** required for the reagent-controlled aldol reaction (Scheme 4). Treatment of aldehyde **13** with the tin enolate derived from oxazolidine-2-thione **14**^{9,17} afforded the desired *syn*-propionate **15**, which when exposed to NaBH_4 gave the diol **16** as a result of reductive auxiliary cleavage.

Removal of the C18 TBS ether required warming to 50 °C to proceed at an appreciable rate, and selective primary/secondary alcohol protection yielded bis-TBS ether **17** (Scheme 5). The stage was now set for the somewhat risky

Scheme 5



late-stage introduction of the C18 hemisuccinate. Several methods for the acylation of hindered alcohols failed,¹⁸ affording mostly the isomerized 5,6-spiroketal as found in **2**, so we turned to the application of high pressure according to the procedure described by Shimizu and Nakata.^{3,19} Thus, treatment of a mixture of alcohol **17** and monoprotected succinic acid **18**^{9b} with DCC and DMAP at 0.4 GPa in CH_2Cl_2 solvent gave the desired ester **19** in very high yield. Interestingly, we found that the reaction proceeded efficiently at much lower pressures than reported previously (1.5 GPa)^{3,19} and required the use of relatively simple high-pressure equipment. Selective deprotection of the primary alcohol in **19** proceeded well using $\text{HF}\cdot\text{pyridine}$ complex buffered with pyridine²⁰ to afford the alcohol **20**. Hydrostannylation of the alkyne in **20** proved somewhat difficult under Pd-catalyzed conditions²¹ owing to the steric hindrance

(15) Dess, D. B.; Martin, J. C. *J. Am. Chem. Soc.* **1991**, *113*, 7277.

(16) Müller, S.; Liepold, B.; Roth, G. J.; Bestmann, J. *Synlett* **1996**, 521.

(17) Nagao, Y.; Hagiwara, Y.; Kumagi, T.; Ochiai, M.; Inoue, T.; Hashimoto, K.; Fujita, E. *J. Org. Chem.* **1986**, *51*, 2391.

(18) For example see: Zhao, H.; Pendri, A.; Greenwald, R. B. *J. Org. Chem.* **1998**, *63*, 7559.

(19) Shimizu, T.; Hiramoto, K.; Nakata, T. *Synthesis* **2001**, 1027.

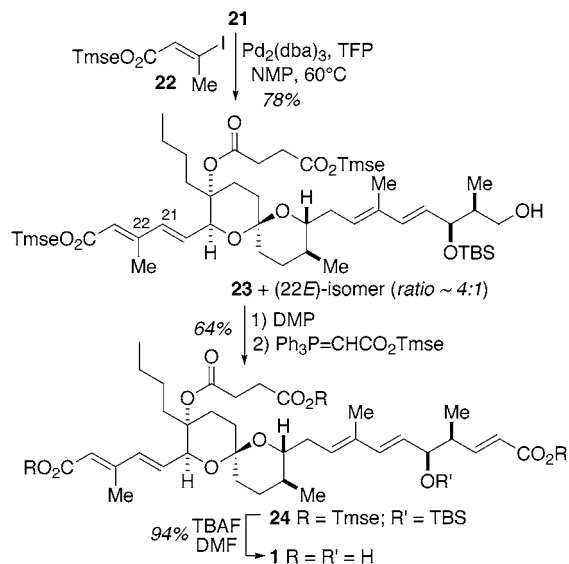
(20) Evans, D. A.; Gage, J. R.; Leighton, J. L. *J. Am. Chem. Soc.* **1992**, *114*, 9434.

(21) Zhang, H. X.; Guibé, F.; Balavoine, G. *J. Org. Chem.* **1990**, *55*, 1857.

at the axial alkyne functionality, and excessive amounts of protodestannylation occurred during the reaction. We found that slow addition of the catalyst (up to 30 mol % over 5 h) was required to give the vinyl stannane **21** in acceptable yields.

The final steps to the target **1** are outlined in Scheme 6. Stille coupling between stannane **21** and vinyl iodide **22**^{9b} afforded the required tetraene **23** along with a small amount of what was tentatively identified as the 22*E*-isomer. This type of isomerization was observed by us during our studies on the total synthesis of reveromycin B (**2**).⁹ Oxidation of

Scheme 6



the free primary alcohol and Wittig extension²² afforded the fully protected reveromycin A precursor **24**, which upon exposure to TBAF in DMF gave reveromycin A (**1**) in high yield. The synthetic material was purified by reverse-phase chromatography (C18 SPE cartridge, 900 mg, 40–60% MeOH/H₂O) and was identical (NMR, IR, UV, HRMS) to the natural product in all respects $\{[\alpha]^{22}_{\text{D}} -122.4$ (c, 0.16, MeOH); lit.^{1b} $[\alpha]^{23}_{\text{D}} -115$ (c, 0.1, MeOH)} including retention time on RP-HPLC.

In conclusion, we have completed the total synthesis of (–)-reveromycin A (**1**), which utilized a HDA reaction hydroboration sequence to construct the spiroketal core in a convergent and stereoselective manner. Application of this methodology to related natural products is underway.²³

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Supporting Information Available: Characterization data for compounds **1**, **3**, **4**, **9–12**, **15–17**, **19**, **20**, **23**, and **24** and copies of the NMR spectra, as well as a comparison of ¹³C NMR chemical shifts and HPLC traces for natural and synthetic reveromycin A (**1**). This material is available free of charge via the Internet at <http://pubs.acs.org>.

OL048811L

(22) Hungerbühler, E.; Seebach, D.; Wasmuth, D. *Helv. Chim. Acta* **1981**, *64*, 1467.

(23) Zanatta, S. D.; White, J. M.; Rizzacasa, M. A. *Org. Lett.* **2004**, *6*, 1041.